Assessing and maximizing data quality in macromolecular crystallography

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The quality of macromolecular crystal structures depends, in part, on the quality and quantity of the data used to produce them. Here, we review recent shifts in our understanding of how to use data quality indicators to select a high resolution cutoff that leads to the best model, and of the potential to greatly increase data quality through the merging of multiple measurements from multiple passes of single crystals or from multiple crystals. Key factors supporting this shift are the introduction of more robust correlation coefficient based indicators of the precision of merged data sets as well as the recognition of the substantial useful information present in extensive amounts of data once considered too weak to be of value.

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Introduction

Recent years have seen changes in our understanding of the factors influencing macromolecular crystallographic data quality and in the recommendations for obtaining the highest quality data and selecting an optimal high resolution cutoff for crystallographic refinement. Here, we will focus on three topics related to these changes. First, we discuss the common data quality indicators and their utility. Second, we describe recent results illustrating how high multiplicity[1] can improve data quality.

Third, we review recent reports providing evidence that extending resolution limits beyond conventional cutoffs to include weaker high resolution data can improve phasing results, electron density maps, and refined models. Understanding of these aspects of data quality is critically important because the observed diffraction data are typically the sole source of experimental information available for supporting a crystallographic structure determination. Strategic considerations regarding other aspects of data collection (e.g. [1,2]) and data reduction (e.g. [3,4[••]]) are also important, but are beyond the scope of this review.

Common data quality indicators

In Table 1, we list and comment on the utility of eight common statistical indicators reported by current data reduction software, including the new $CC_{1/2}$ and CC^* [5[••]]. The equations for each are in the literature and are not given here. These indicators all report on data *precision*, so if substantial systematic errors are present the indicators need not reflect the data *accuracy* [4[••]]. We have arranged the data precision indicators into three groups according our view of their utility, and we also specify for each one the crucial distinction of whether it reports on the precision of *individual* or of *merged* measurements (Table 1).

With the introduction of $CC_{1/2}$, all three key indicators we primarily recommend for assessing the precision of the merged data (for both standard and serial crystallography) are Pearson's correlation coefficients (CC) between independent sets of observations characterized as a function of resolution: $CC_{1/2}$, $CC_{1/2\text{-anom}}$, and CC^* (Table 1). CC values range from 1 to −1 for perfectly correlated versus anticorrelated data, but for properly indexed data these indicators should range from near 1 for highly precise data to near 0 for very imprecise data. An advantage of CC-based indicators is that they have well-studied statistical properties so that, for instance, given a CC value and how many observations contributed to it, one can calculate the probability that this value has occurred by chance, i.e. how likely it is that the null hypothesis holds (e.g. http://en.wikipedia.org/wiki/Statistical_significance). $CC_{1/2\text{-anom}}$, our suggested name for the correlation between independent estimates of the anomalous differences from half data sets [6], was the first of these CC-based indicators to be introduced as an extension of work showing that the CC between anomalous differences of two complete data sets helped define which data would be useful for solving anomalous substructures [7].

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[1] We prefer the term multiplicity to the more common redundancy as it emphasizes that the multiple observations of a reflection are not actually redundant with each other, but together provide more information than any of the individual observations.

Table 1

Common indicators of data precision and their recommended usage. The 'type' defines whether an indicator reports on the precision of individual observations or the final merged data

Indicator	Type	Recommended usage
Indicators of first rank		
$CC_{1/2}$	Merged	CC between intensity estimates from half data sets. Primary indicator for use for selecting high resolution cutoff for data processing. Is related to the effective signal to noise of the data (see Box 1).
$CC_{1/2\text{-anom}}$	Merged	Suggested shorthand for CC between anomalous difference estimates from half data sets. (We suggest CC_{anom} be used if datasets rather than between half data sets are compared [7].) Primary indicator for assessing the resolution limit of useful anomalous signal. Analogous to CC*, a CC^*_{anom} indicator can be calculated from $CC_{1/2\text{-anom}}$.
CC*	Merged	Calculated from $CC_{1/2}$. Indicator useful for comparing data and model quality. Provides the potential for a cross-validation independent indication of overfitting [5••]. CC* is undefined for negative $CC_{1/2}$.
Additional useful indicators		
R_{meas} (=R_{rim})	Individual	Multiplicity independent replacement of R_{merge} and R_{sym}; Useful for assessing space group symmetry and isomorphism of multiple data sets; Should play no role in determining resolution cutoff.
R_{pim} ($\sim R_{mrgdI}/\sqrt{2}$)	Merged	Mainly of value for comparisons with previous practices; Should play no role in determining resolution cutoff, as it rises toward infinity as signal decreases; Also because it is an unweighted sum, if data of varying quality are merged, it will underestimate the quality of the final data. The SFX community's $R_{split} = R_{mrgdI}$.
$\langle I/\sigma \rangle_{ind}$	Individual	Average signal-to-noise ratio of individual observations. The σ for each reflection is calculated according to an 'error model' that parameterizes the random and systematic errors. Should play no role in determining resolution cutoff.
$\langle I/\sigma \rangle_{mrgd}$	Merged	As $\langle I/\sigma \rangle_{ind}$ but for the intensities after a weighted averaging ('merging') of equivalent observations. For reflections with multiplicity n, $\langle I/\sigma \rangle_{mrgd}$ is at most $\sqrt{n}$ higher than $\langle I/\sigma \rangle_{ind}$, but the increase will be less if reflections to be merged have varying $\langle I/\sigma \rangle_{ind}$. More useful than $CC_{1/2}$ for assessing quality of low resolution data. If properly estimated data should correlate with $CC_{1/2}$ values (see Box 1).
ISa	Individual	Also $\langle I/\sigma \rangle_{asymptotic}$; theoretical value of $\langle I/\sigma \rangle_{ind}$ for an infinitely strong observation of the dataset calculated from coefficients of error model established during scaling [3,17]. Gives insight into the level of fractional error in the dataset.
Indicators that should not be used		
$R_{merge} = R_{sym}$	Individual	Flawed indicators that have been replaced by R_{meas}. We recommend these be removed from all data reduction software.
Overall value	Both	'Overall' quantities for statistics are not of general value, because they are highly influenced by the distribution of multiplicity. More informative would be reporting 'low resolution bin' and 'high resolution bin' values.

Similarly $CC_{1/2}$, calculated in resolution shells by correlating the intensity values produced from two half data sets [5••], provides a model-free, empirical measure of the level of discernable signal and is equivalent to the Fourier Shell Correlation statistic used to define resolution in cryo-EM studies (e.g. [8]). In fact, a theoretical relationship between $CC_{1/2}$ and the signal-to-noise of the merged data ($\langle I/\sigma \rangle_{mrgd}$) can be derived that helps put $CC_{1/2}$ on a familiar footing (Box 1). Typically, $CC_{1/2}$ is near 1.0 (or 100%) at low resolution, and drops smoothly toward 0 as the signal-to-noise ratio decreases. Any deviations from this behavior should be scrutinized as possible indicators of anomalies. Since $CC_{1/2}$ measures how well one half of the data predicts the other half, it does not directly indicate the quality of the data set after final merging. This, however, is estimated by the quantity CC*.

CC* is mathematically derived from $CC_{1/2}$ using the relationship CC* = $[2\,CC_{1/2}/(1 + CC_{1/2})]^{1/2}$ and provides an estimate of the CC that would be obtained between the final merged data set and the unknown true values that they are representing [5••]. This brings a new ability

to compare data and model quality on the same scale because one can compare CC* with a CC between F^2_{calc} and F^2_{obs} (i.e. CC_{work} or CC_{free}) to discover even without cross-validation if overfitting has occurred during refinement [5••]. The calculations have been built into the PHENIX system [9], and already been used in some reports (e.g. [10•,11••]). While how to best use this information in guiding and validating refinements is not yet clear, it provides a welcome replacement for the practice of comparing refinement R-factors with data reduction R-factors (e.g. [12•]) that is not correct [5••].

In our view (Table 1), no indicators other than $CC_{1/2}$ should influence the high-resolution cutoff decisions for data processing. As noted (Box 1), the $\langle I/\sigma \rangle_{mrgd}$ statistic (we use subscripts for the two different $\langle I/\sigma \rangle$ values to avoid confusion and emphasize their distinct information content) is related to $CC_{1/2}$, and so is in principle equally useful for defining a cutoff. However, it is not as useful in practice because the $\langle I/\sigma \rangle_{mrgd}$ values obtained during data reduction may not be accurate since they depend on the error model and parameterization used and additional

Box 1 Approximate relation between $CC_{1/2}$ and $\langle I/\sigma \rangle_{mrgd}$. Assuming that the $\sigma = \sigma_{mrgd}$ values obtained from data processing are consistent with the spread of observations around their mean, we can derive an approximate expected relationship between $CC_{1/2}$ and $\langle I/\sigma \rangle_{mrgd}$ in a high resolution shell. From the derivation of equation 1 in Karplus and Diederichs [5] we have

$$CC_{1/2} = \frac{\sigma_\tau^2}{\sigma_\tau^2 + \sigma_\epsilon^2} = \frac{<I^2> - <I>^2}{<I^2> - <I>^2 + \sigma_\epsilon^2}$$

where σ_ϵ denotes the mean error within a half-dataset. Introducing $q^2 = \langle I^2 \rangle / \langle I \rangle^2 - \langle I \rangle^2$, we can write

$$CC_{1/2} = \frac{1}{(1 + q^2 \sigma_\epsilon^2 / \langle I \rangle^2)}$$

At this point, we note that for acentric reflections following a Wilson distribution $q^2 = 2$ and $\sigma_\epsilon^2 = 2\langle \sigma \rangle^2$, which lets us write $CC_{1/2}$ for acentric reflections as

$$CC_{1/2}^{acentric} = \frac{1}{[1 + 4/(\langle I \rangle / \langle \sigma \rangle)^2]}$$

Then, since $\langle I/\sigma \rangle$ is close to $\langle I \rangle / \langle \sigma \rangle$, in particular at high resolution where σ is approximately the same for all reflections, it is a reasonable approximation that

$$CC_{1/2} \sim \frac{1}{[1 + 4/\langle I/\sigma \rangle_{mrgd}^2]}$$

Two factors that may shift this relationship are (1) that real data may include some centric reflections, for which $q^2 = 3/2$, changing the 4 in the above equations to a 3, and (2) that at very low $\langle I/\sigma \rangle_{mrgd}$ the measured intensities are dominated by Gaussian noise and will not follow Wilson statistics and $q^2 = 1$ applies, which changes the 4 in the above equations to a 2. Thus in resolution shells having weak data, the $CC_{1/2}$ versus $\langle I/\sigma \rangle_{mrgd}$ relationship should be fall between the extreme cases of:

$$CC_{1/2} \sim \frac{1}{[1 + 4/\langle I/\sigma \rangle_{mrgd}^2]} \quad \text{and} \quad CC_{1/2} \sim \frac{1}{[1 + 2/\langle I/\sigma \rangle_{mrgd}^2]}$$

The relationship should tend to be closer to the first equation for data with $\langle I/\sigma \rangle_{mrgd}^2 \geq 1$ for which Wilson statistics are still relevant. As seen in the figure, this implies that for accurately estimated σ_{mrgd} values, $CC_{1/2}$ between ~ 0.1 and ~ 0.4 can be roughly equated to $\langle I/\sigma \rangle_{mrgd}$ values between ~ 0.5 and ~ 1.5.

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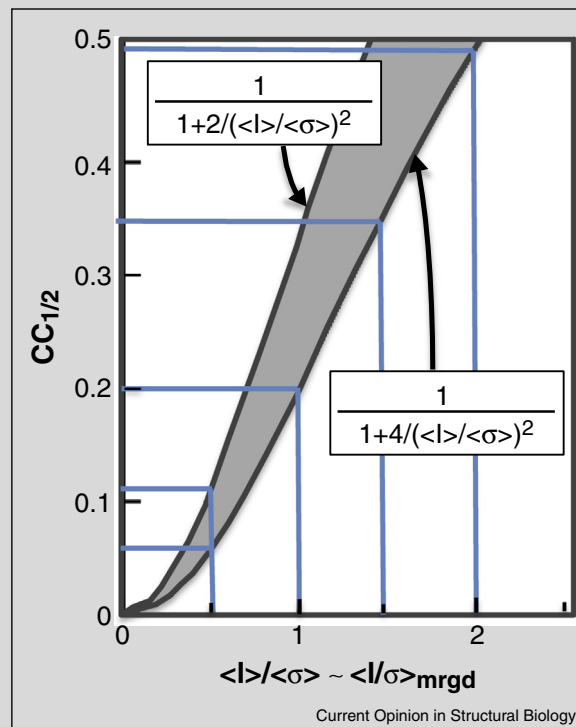
Figure legend: The two curves define limiting relationships for how $CC_{1/2}$ relates to $\langle I \rangle / \langle \sigma \rangle$ for low signal data. The upper curve is only valid if Gaussian noise dominates the data; so the lower curve should be considered the more relevant above $\langle I \rangle / \langle \sigma \rangle \sim 0.5$. The pale blue lines highlight the corresponding values for $\langle I \rangle / \langle \sigma \rangle$ of 0.5, 1.0, 1.5 and 2.0.

factors such as outlier rejection algorithms and weighting of observations. This causes some irreproducibility across programs as was documented in a report showing that data processed by HKL2000, MOSFLM, and XDS yielded at a certain resolution $\langle I/\sigma \rangle_{mrgd}$ values of 2.7, 3.5, and 5.2, respectively [13].

To illustrate why the other common indicators (besides $CC_{1/2}$ and $\langle I/\sigma \rangle_{mrgd}$) are not useful for guiding the high resolution cutoff decision, we offer the following gedanken experiment. Consider five idealized datasets without radiation damage or systematic errors: '*Big*' with multiplicity = 2 from a rare large crystal; '*Tiny*' from a readily grown 100-fold smaller microcrystal; '*T100*' resulting from the merging of 100 equivalent microcrystal datasets; '*Big+T100*' resulting from the merging of *Big* and *T100*; and '*Big2*' resulting from the merging of *Big* with an equivalent dataset from a second large crystal. Assuming $\langle I/\sigma \rangle_{ind} = 2$ in the highest resolution bin of *Big*,

and that $R_{meas} \sim 0.8/(\langle I/\sigma \rangle_{ind})$ [4$^{\bullet\bullet}$], that $CC_{1/2}$ is related to $\langle I/\sigma \rangle_{mrgd}$ as shown in Box 1, and that n-fold repetition of a measurement reduces its σ by $\sqrt{n}$, we can generate the following idealized high resolution bin statistics for the five datasets:

Dataset	Big	Tiny	T100	Big+T100	Big2
Multiplicity	2	2	200	202	4
$\langle I/\sigma \rangle_{ind}$	2.0	0.2	0.2	0.22	2.0
R_{merge}	28%	280%	399%	395%	35%
R_{meas}	40%	400%	400%	396%	40%
R_{pim}	28%	280%	28%	28%	20%
$\langle I/\sigma \rangle_{mrgd}$	2.8	0.28	2.8	4.0	4.0
$CC_{1/2}$	0.66	0.04	0.66	0.80	0.80

Readily apparent is that according to $CC_{1/2}$ and $\langle I/\sigma \rangle_{mrgd}$, *Big* and *T100* are of equivalent quality as are *Big+T100* and *Big2*. The huge differences in the values of $\langle I/\sigma \rangle_{ind}$, R_{merge}, and R_{meas} within these pairs shows why indicators of the precision of individual measurements should *never*

be used for guiding cutoff decisions. Furthermore, a comparison of *Big+T100* vs. *Big2* shows that even R_{pim} does not reflect their equivalence. This is because when data of different precision are merged, R_{pim} and all R-factor based indicators lose relevance because each reflection is weighted equally rather than (as for $\langle I/\sigma \rangle_{mrgd}$) according to its reliability. The impact of this is even more dramatically seen in the 10-fold different R_{meas} values of *Big+T100* vs. *Big2*. Also worth noting is that *T100* has quite respectable signal in the highest resolution bin even though *Tiny* does not, emphasizing that high resolution cutoff decisions should only be made *after* all relevant data have been merged. Finally, the large increase in R_{merge} for *Tiny* vs. *T100* reveals how tremendously misleading is the overestimation by R_{merge} of precision at low multiplicity [14,15] — making it appear that the datasets merged in *T100* were not isomorphous and should not be merged, even while R_{meas} correctly indicates the merged data were isomorphous. This is why we recommend (Table 1) that R_{merge} never be used.

Further, we suggest that publication standards be changed to require low and high resolution shell data quality statistics rather than 'overall' and high resolution shell values (Table 1). It has been shown that 'overall' statistics are weighted by multiplicity [14], and so depending on how multiplicity varies with resolution, the 'overall' number can take on any value from that of the strongest data to that of the weakest data. Selecting a more generous high resolution cutoff (e.g. [11••,16•]) and/or increasing the multiplicity of the high resolution data (e.g. [14]), makes the 'overall' statistics become worse even though the resulting data are better.

Finally, one uncommon indicator we recommend be reported by data reduction programs (but not in structure reports) is $\langle I/\sigma \rangle_{asymptotic}$ or 'ISa' ([17]; Table 1). Importantly, 1/ISa provides an estimate of the level of experiment/hardware related systematic (i.e. fractional) error in the data set that limits the precision of strong reflections. For instance, an ISa of near 30, about as high as can be achieved for CCD detector data sets [4••], indicates about a 3.3% (i.e. ~1/30) systematic error. ISa thus has utility as a diagnostic for guiding efforts to improve experimental setups as well as data processing.

Multiplicity can powerfully enhance measurable signal through decreasing noise
A crucial distinction to make regarding data quality is the difference between the level of signal that is measured in a particular data set versus the level of signal that could in principle be measured from that sample (e.g. Figure 1a). Given only random errors, the standard error σ in a measurement is reduced by $\sqrt{n}$ if the measurement is repeated n times. The utility of high multiplicity data sets from single crystals to improve the accuracy of anomalous signal measurements and enable phasing has been powerfully demonstrated many times (e.g.

[18,19,20,21]), and it has been recognized that for success sufficient data must be collected before radiation decay degrades the signal (e.g. Figure 1a; [22,23]). Furthermore, theory and practice agree that for a given total crystal exposure time, fractional errors associated with data collection can be minimized and better data produced by collecting higher multiplicity data using shorter exposures [24].

An important recent advance has been the (re)-discovery that high multiplicity can improve signal strength, even for anomalous signal, through combining of data from multiple crystals [25••,26•,27] as long as the individual data sets are tested for isomorphism. Building on this work, Akey *et al.* ([11••]) merged data from 18 crystals to generate anomalous data with 50-fold multiplicity and data for refinement with 100-fold multiplicity thus enabling phasing and a higher resolution refinement than could be accomplished using data from any single crystal. These data illustrate how during merging, the final R_{meas} becomes roughly the average of the individual R_{meas} values, but the final $\langle I/\sigma \rangle_{mrgd}$ and $CC_{1/2}$ values can improve substantially (Figure 1b–d). Recent serial femtosecond crystallography (SFX) results, for which each crystal only provides a single image [28], provide further examples of the power of enhancing data quality through merging data from multiple crystals. In a time-resolved SFX study of the photoactive yellow protein, to obtain sufficient quality difference electron density maps, the workers aimed for a multiplicity of ~1500 in the highest resolution bin [29•].

Evidences that data beyond $R_{meas} \sim 60\%$ and $\langle I/\sigma \rangle_{mrgd} \sim 2$ contain useful information
Until recently, a common and recommended practice has been truncating data at the resolution at which R_{meas} remains below ~60% and $\langle I/\sigma \rangle_{mrgd}$ is ~2 or higher ([30] and Figure S1 of [5••]). Our report [5••] introducing $CC_{1/2}$ also introduced paired refinement tests and showed that, for our test cases, including data out to a $CC_{1/2}$ value of between 0.1 and 0.2 led to an improved refined model even though the data at that resolution had $R_{meas} \sim 450\%$ and $\langle I/\sigma \rangle_{mrgd} \sim 0.3$. We also showed that these weak data improved the quality of difference maps (see Figure S2 of [5••]). This reinforced earlier evidence for the value in refinement of data having $\langle I/\sigma \rangle_{mrgd} \sim 0.5$ [31,32]. The damage caused by using an $R_{meas} \sim 60\%$ cutoff criterion grows with increasing multiplicity, because the excluded data have a higher and higher $\langle I/\sigma \rangle_{mrgd}$. For instance, for the 100-fold multiplicity data set of Akey *et al.* [11••], an $R_{meas} \sim 60\%$ cutoff corresponds to ~3.7 Å resolution at which $\langle I/\sigma \rangle_{mrgd}$ is ~12 (Figure 2c and d). In another study, extending the resolution from 2.85 Å ($R_{meas} \sim 60\%$) to 2.1 Å ($R_{meas} \sim 680\%$; $CC_{1/2} = 0.22$, $\langle I/\sigma \rangle_{mrgd} = 0.9$) improved the MR-Rosetta [33] solution to a challenging molecular replacement problem from $R_{free} \sim 40\%$ to $R_{free} \sim 31\%$ [34].

In terms of the value of using data beyond $\langle I/\sigma \rangle_{mrgd} \sim 2$, one set of systematic refinement tests showed small

Figure 1
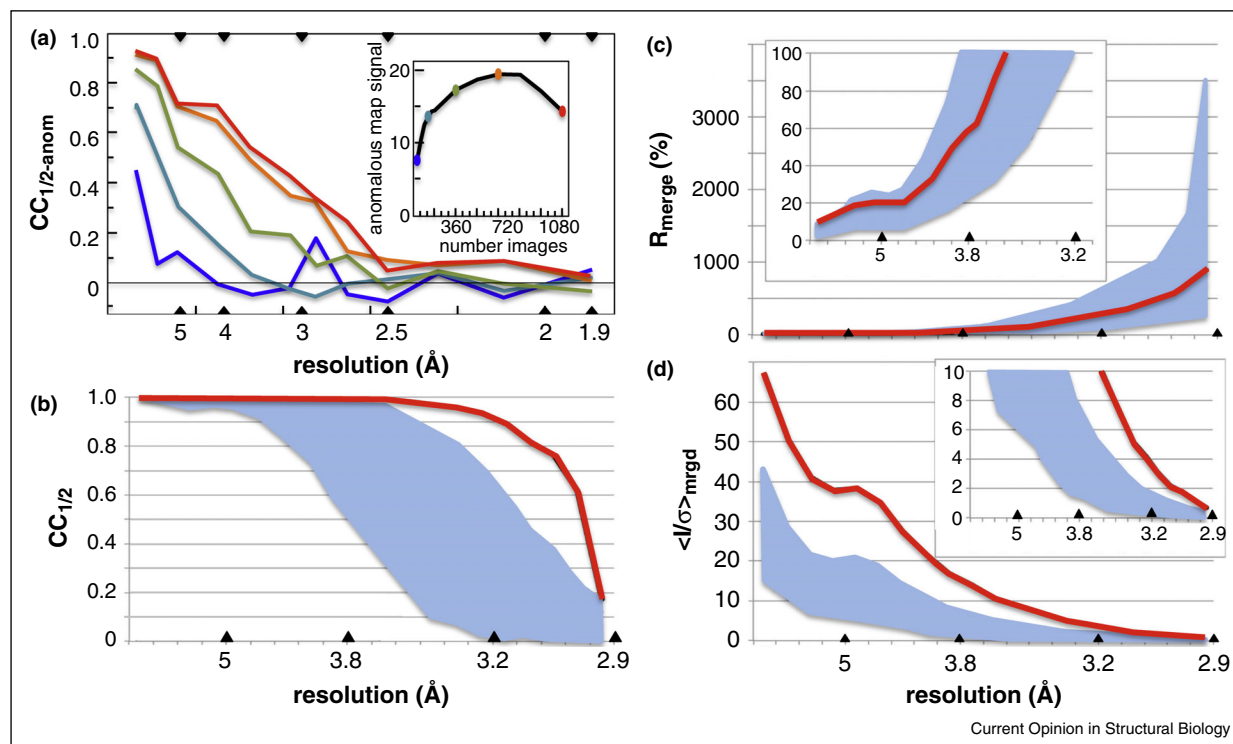
Averaging multiple measurements can substantially enhance data quality. **(a)** CC_{anom} is plotted as a function of resolution for a data set of 1080 $1°$ images in a sulfur-SAD phasing case study [23]. Statistics for data merged from 30 (blue), 120 (cyan), 360 (green), 720 (orange), and 1080 (red) images are shown. Based on 30 images (3.5 fold multiplicity), there is no apparent anomalous signal beyond 4 Å, but with 720 images (75-fold multiplicity) the apparent signal extends beyond 3 Å resolution. Inset shows the quality of the anomalous difference map (maximal r_{rms}) increases substantially and then, as radiation damage systematically alters the structure, decreases even while $CC_{1/2-anom}$ stays high. **(b–d)** Behavior of $CC_{1/2}$, R_{merge} (shown here as the a surrogate to qualitatively reflect the behavior of R_{meas}), and $<I/\sigma>_{mrgd}$ as a function of resolution for individual crystals (breadth of values indicated by cyan swaths) and for a set of data merged from 18 crystals (red traces) and successfully used for sulfur-SAD phasing and refinement at 2.9 Å resolution [11$^{••}$]. Insets show close-ups of the low or high resolution regions. According to the authors, the best individual crystal would only have been useful to ca. 3.2 Å resolution, and by the panel C inset, the averaged data would have been truncated at near 3.7 Å based on an $R_{merge} \sim 60\%$ cutoff criterion.

improvements with no negative impacts by including data out to $CC_{1/2}$ between 0.2 and 0.4 corresponding to $\langle I/\sigma \rangle_{mrgd}$ between 0.5 and 1.5 [35$^{••}$]. Interestingly this correspondence between $\langle I/\sigma \rangle_{mrgd}$ and $CC_{1/2}$ roughly matches that expected from theory (Box 1). Another study using distinct tests similarly concluded that useful information is present in reflections out to $CC_{1/2}$ between 0.1 and 0.5, and that extending the resolution by $\sim$0.2 Å beyond an $\langle I/\sigma \rangle_{mrgd} \sim 2$ cutoff provided a marginal benefit and no adverse effects [36$^{••}$]. A third study showed that the practice of selectively removing weak reflections within a given resolution bin introduced systematic errors into the data and leads to worse refined models [37$^{••}$].

Also, many analyses are now using the more generous $CC_{1/2}$-based cutoffs (with high resolution R_{meas} values as high as $\sim$1000% and $\langle I/\sigma \rangle_{mrgd}$ values as low as $\sim$0.3) and authors comment on the benefits (e.g. [11$^{••}$,16$^{•}$,38,39$^{•}$]). One striking example is shown in Figure 2a. Weak data have further been shown to improve the phasing of

a crystal with 16-fold non-crystallographic symmetry. Phase extension and automated modeling using data truncated per conventional criteria at 3.1 Å resolution stalled at $R_{free} \sim 35\%$, whereas using an extended 2.5 Å resolution cutoff produced an excellent model with $R_{free} \sim 24.5\%$ and improved electron density maps (Figure 2b; [40$^{••}$]). Although the signal per reflection is rather weak for the extended data, the tangible impact on phase extension, refinement, and map quality can be rationalized in that the numbers of added reflections are very large – in some cases doubling the data available – and they help to minimize series termination error. Wang [41$^{•}$] describes a perverse incentive that may tempt researchers to truncate datasets to obtain more attractive R/R_{free} values for any given model, and proposes an intriguing modified Rfactor that emphasizes the value of using more data.

As there is no single 'correct' cutoff for every case, using paired refinements [5$^{••}$] provides a controlled approach

Figure 2
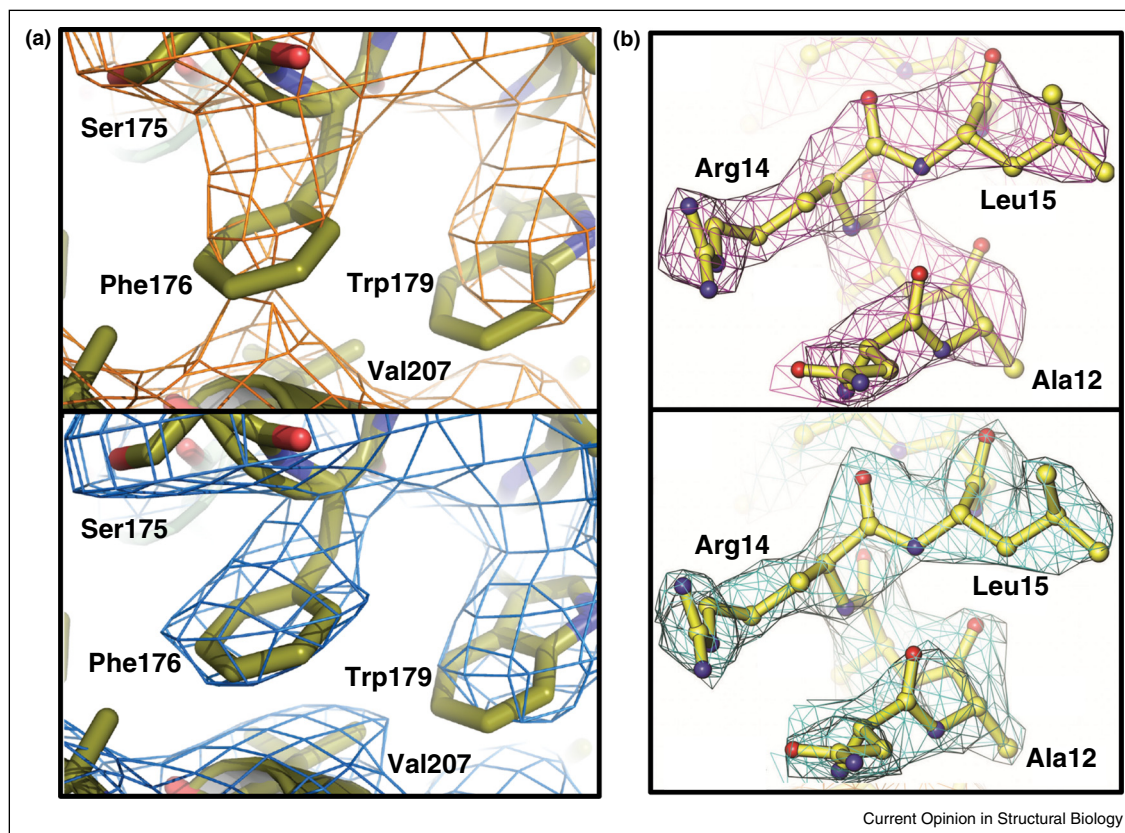
Examples of tangible electron density map improvement enabled by extending resolution cutoffs. **(a)** Comparison of the 2Fo–Fc electron density (contoured at 1 ρ_{rms}) for a region of the prokaryotic sodium channel pore using an $\langle I/\sigma \rangle_{mrgd} \sim 2$ cutoff (R_{pim} = 47%, $\langle I/\sigma \rangle_{mrgd}$ = 1.9, $CC_{1/2}$ = 0.78) of 4.0 Å resolution (upper panel) versus a more generous $CC_{1/2} \sim 0.1$ based cutoff (R_{pim} = 213%, $\langle I/\sigma \rangle_{mrgd}$ = 0.3, $CC_{1/2}$ = 0.14) of 3.46 Å resolution (lower panel). The 4 Å resolution cutoff was already somewhat generous as the R_{pim} of 47% with a multiplicity of 12 would be expected to correspond to an R_{meas} value of above 150% (47%*$\sqrt{12}$). Used with permission from Figure S1 of [48●]. **(b).** Comparison of the 2Fo–Fc electron density (contoured at 1 ρ_{rms}) for a region of the *E. coli* YfbU protein using for the phase extension a fairly conventional cutoff (R_{meas} = 77%, $\langle I/\sigma \rangle_{mrgd}$ = 3.5, $CC_{1/2}$ = 0.85) of 3.1 Å resolution (upper panel) versus a more generous $\langle I/\sigma \rangle_{mrgd} \sim 0.5$ or $CC_{1/2} \sim 0.1$ cutoff (R_{meas} = 302%, $\langle I/\sigma \rangle_{mrgd}$ = 0.5, $CC_{1/2}$ = 0.14) of 2.5 Å resolution (lower panel). The additional weak data did not just extend the resolution of the map, but improved the quality of the phases obtained at 3.1 Å resolution. Images used with permission from the International Union of Crystallography from Figure 3 of [40●●] (http://dx.doi.org/10.1107/S1399004714005318).

to decide for any dataset what resolution cutoff yields the best model. And the PDB-REDO server is now available as a refinement tool that includes a paired refinement option [42●]. Another conservative approach to the cutoff question is to process one's data out to $CC_{1/2} \sim 0.1$, but carry out initial refinements using a self-selected conservative resolution limit until the residual Fo–Fc difference map has no interpretable peaks. Then, one can recalculate the difference map using an extended resolution cutoff, and any interpretable peaks provide evidence of tangible information brought by the newly included weak data. In one project for which this was done, the extended difference map was highly informative, and further refinement improved our 2.6 Å resolution 'final' model with R/R_{free} = 18.9/23.2% to a lower R/R_{free} = 17.4/22.0% even at the extended 2.3 Å resolution [43●].

Conclusions and outlook

When we pointed out the flawed multiplicity dependence of R_{merge} and recommended making resolution cutoff decisions based on precision of the data *after* merging [14], we predicted that this 'should stimulate a shift in data collection strategies, so that the current bias toward using single crystals for complete data sets whenever possible will shift to favor multiple crystal data sets which have increased multiplicity and hence more accurate reduced structure factors.' The continued use of R_{merge} (or even R_{meas}) to define cutoffs hindered this from occurring, but now, with the introduction of $CC_{1/2}$ as a statistically robust indicator of the precision of merged data and with definitive evidence that the inclusion of weak data improves models and that merging data from multiple crystals can be highly beneficial, practices are changing in this direction. Increasing multiplicity is a

reasonable strategy to pursue not just to enhance anomalous signals for phasing, but also for obtaining the best high resolution data set for refinement. Important to note, though, is increasing the signal-to-noise of high resolution data occurs by decreasing noise rather than increasing the intensity, so it does not increase the relative contribution of these structure factors to the electron density map. Also worth noting is that obtaining the best high resolution data does not guarantee the best model will be obtained; that depends on care being taken by the crystallographer during model building and refinement.

Further, there is a clear need for changes to what journals require for Table 1 statistics, and we think a useful set would simply be the high resolution bin $CC_{1/2}$ and number of reflections it is based on (since the low resolution bin $CC_{1/2}$ is always ~1 and not very informative) and $\langle I/\sigma \rangle_{mrgd}$ in the low and high resolution bins, and potentially (for calibration with the past) the resolution at which $\langle I/\sigma \rangle_{mrgd} \sim 2$. $CC_{1/2}$ also needs to be added to the PDB deposition form. In terms of what this does to the meaning of resolution, we are in agreement with Phil Evans [44] that the nominal resolution of a structure has always referred to which reflections are included in the Fourier summation rather than guaranteeing a certain quality in terms of the apparent resolution of the resulting electron density maps.

We also support the ongoing efforts to archive raw diffraction data to maximize the potential benefit of research funds invested by providing maximal flexibility for correcting mistakes and improving existing structures as technologies improve [45–47]. In the meantime, we encourage users to process data out to $CC_{1/2} \sim 0.1$ (after merging of crystals!) even if one is not planning to use it, and to deposit *unmerged* intensity values together with the merged values. While much is still to be learned about how to obtain the best data, we hope the examples provided here will help crystallographers collect better data and determine better structures.

Conflict of interest statement

The authors have no conflicts of interest in preparing this work.

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Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

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•• *Nucleic Acids Crystallography: Methods and Protocols*, vol. 320. Edited by Ennifar . New York: Springer Science + Business Media; 2015 (in press).
Provides a rather in-depth discussion of random and systematic errors that impact crystallographic data quality, the distinction between precision and accuracy, and the use of data quality indicators. Then walks readers through some lesser known tools that can be used to troubleshoot and optimize data reduction using XDS.

5. Karplus PA, Diederichs K: **Linking crystallographic model and**
•• **data quality**. *Science* 2012, **336**:1030-1033.
Introduces paired refinement concept, $CC_{1/2}$ and CC^* indicators. Uses these and difference map analyses to prove that conventional high-resolution cutoff criteria discard useful data. Further shows how data quality indicators are not comparable to refinement R-factors and that their values should not be used in defining a high-resolution cutoff. That the new indicators are being used and practices are changing is indicated by the over 400 citations already garnered by the work.

6. Evans P: **Scaling and assessment of data quality**. *Acta Crystallogr D: Biol Crystallogr* 2006, **62(Pt 1)**:72-82.

7. Schneider TR, Sheldrick GM: **Substructure solution with SHELXD**. *Acta Crystallogr D: Biol Crystallogr* 2002, **58(Pt 10 Pt 2)**:1772-1779.

8. Scheres SH, Chen S: **Prevention of overfitting in cryo-EM structure determination**. *Nat Methods* 2012, **9**:853-854.

9. Adams PD, Afonine PV, Bunkoczi G, Chen VB, Davis IW, Echols N, Headd JJ, Hung LW, Kapral GJ, Grosse-Kunstleve RW, McCoy AJ *et al.*: **Phenix: a comprehensive python-based system for macromolecular structure solution**. *Acta Crystallogr D: Biol Crystallogr* 2010, **66(Pt 2)**:213-221.

10. Bae B, Davis E, Brown D, Campbell EA, Wigneshweraraj S,
• Darst SA: **Phage T7 Gp2 inhibition of *Escherichia coli* RNA polymerase involves misappropriation of σ^{70} domain 1.1**. *Proc Natl Acad Sci U S A* 2013, **110**:19772-19777.
Early example of low resolution (~4 Å) structure analyses using extended $CC_{1/2}$-based resolution limits and including supplementary plots showing CC_{work} and CC_{free} behavior along with R_{work} and R_{free} showing that despite the use of data with, in one case, R_{meas} values over 400% the refinements behave well.

11. Akey DL, Brown WC, Konwerski JR, Ogata CM, Smith JL: **Use**
•• **of massively multiple merged data for low-resolution S-SAD phasing and refinement of flavivirus ns1**. *Acta Crystallogr D: Biol Crystallogr* 2014, **70(Pt 10)**:2719-2729.
Along with the *de novo* solution of a large protein structure by S-SAD phasing enabled by merging data from many crystals, the authors provide a retrospective analysis of how the high multiplicity improved the data, how excluding data from certain crystals improved the results, and how using data well-beyond conventional resolution limits immensely improved the structure determination.

12. Holton JM, Classen S, Frankel KA, Tainer JA: **The R-factor gap in**
• **macromolecular crystallography: an untapped potential for insights on accurate structures**. *FEBS J* 2014, **281**:4046-4060.
A valuable study using real and simulated data to show that substantial information exists in diffraction data that are not accounted for by refined protein models, and suggests these are related to inadequately modeled aspects of protein dynamics and the solvent organization. Relevant to this review is the evidence it provides of the deeply engrained nature of certain misconceptions regarding data quality indicators. These include the use of R_{merge} much more than R_{meas}, the incorrect consideration of these values rather than R_{pim} as being relevant to the quality of the merged data, and finally the treating of intensity-based data quality R-factors of any kind as being quantitatively comparable with the crystallographic R-factors from refinement. We think that the conclusions of the work would still hold up using more appropriate comparators, and may in fact even become stronger.

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14. Diederichs K, Karplus PA: **Improved R-factors for diffraction data analysis in macromolecular crystallography**. *Nat Struct Biol* 1997, **4**:269-275.

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• **Structure and function of human xylulokinase, an enzyme with important roles in carbohydrate metabolism**. *J Biol Chem* 2013, **288**:1643-1652.
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17. Diederichs K: **Quantifying instrument errors in macromolecular X-ray data sets**. *Acta Crystallogr D: Biol Crystallogr* 2010, **66(Pt 6)**:733-740.

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•• Mancia F, Hendrickson WA: **Structures from anomalous diffraction of native biological macromolecules**. *Science* 2012, **336**:1033-1037.
Extension of Liu et al. [27] work to show that with filtering for non-isomorphous crystals, improved anomalous signals achieved by merging the data from multiple crystals to give a multiplicity of 100–150 allows S-SAD phasing for five proteins. Even individual data sets with no apparent signal on their own (i.e. $CC_{1/2-anom} \sim 0$) could be combined to provide a merged data set with substantial signal emphasing that 'no apparent signal present' does not mean 'no signal present.'

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• **native biological macromolecules from multi-crystal anomalous diffraction data**. *Acta Crystallogr D: Biol Crystallogr* 2013, **69(Pt 7)**:1314-1332.
Further details of the multi-crystal native SAD phasing approach reported in [25••], through a focus on one of the five proteins in that study — the DnaK:ATP complex. Includes a discussion of broader aspects of such analyses supporting the conclusion that the appropriate merging of data from multiple statistically equivalent crystals should be generally applicable.

27. Liu Q, Zhang Z, Hendrickson WA: **Multi-crystal anomalous diffraction for low-resolution macromolecular phasing**. *Acta Crystallogr D: Biol Crystallogr* 2011, **67(Pt 1)**:45-59.

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35. Evans PR, Murshudov GN: **How good are my data and what**
•• **is the resolution?** *Acta Crystallogr D: Biol Crystallogr* 2013, **69(Pt 7)**:1204-1214.
Introduces the new scaling program AIMLESS and has an extensive, thoughtful discussion of how to choose a high resolution cutoff. Included are tests comparing the impact on refinement of including real versus fake weak high resolution data that lead to the conclusion that 'by the time $CC_{1/2}$ has fallen to around 0.2–0.4, or $\langle I/\sigma \rangle$ to around 0.5–1.5, there is little information remaining.' Appendix 1 shows that for a refinment against random data, R_{free} approaches 42% rather than a value around 55–60% as is commonly assumed.

36. Luo Z, Rajashankar K, Dauter Z: **Weak data do not make a free**
•• **lunch, only a cheap meal**. *Acta Crystallogr D: Biol Crystallogr* 2014, **70(Pt 2)**:253-260.
Excellent background discussion regarding resolution cutoffs, and careful studies of four 1.4–2 Å resolution test cases. Tests comparing observed vs. randomized weak data imply it is genereally helpful to include data $\sim$0.2 Å beyond an $\langle I/\sigma \rangle_{mrgd} = 2$ cutoff; however, a surveyed set of quantitive model quality indicators (R_{free}, estimated coordinate error, and maximum likelihood target functions) did not show consistent favorable changes in both Refmac and PHENIX refinements. Interestingly, the optical resolution of electron density maps was shown to increase upon the inclusion of even the weakest data tested (in their Figure 3). Despite the minimal tangible evidences of model improvements, the authors concluded that the 'extension of the maximum resolution at the stage of data collection and structure refinement is cheap in terms of the required effort and is definitely more advisable than accepting a too conservative resolution cutoff.'

37. Diederichs K, Karplus PA: **Better models by discarding data?**
•• *Acta Crystallogr D: Biol Crystallogr* 2013, **69(Pt 7)**:1215-1222.
Shows using paired refinements that merging a weaker data set with a stronger one can produce improved models, and that selectively removing weak data from datasets leads to worse models even though data quality R-factors are lower. Also shows that in these tests $CC_{1/2}$ even more than $\langle I/\sigma \rangle_{mrgd}$ predicts which data sets will produce better models systematic error are documented and discussed, presumably because $\langle I/\sigma \rangle_{mrgd}$ can increase even when data quality is not improving, whereas the empirical $CC_{1/2}$ accurately reflects when merging certain data actually degrades the precision. Also includes a theoretical analysis of how some systematic errors (including those introduced by certain data filtering practices) impact $CC_{1/2}$.

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39. Noeske J, Wasserman MR, Terry DS, Altman RB, Blanchard SC,
• Cate JH: **High-resolution structure of the *Escherichia coli* ribosome**. *Nat Struct Mol Biol* 2015, **22**:336-341.
Using a $CC_{1/2}$ significant at $p < 0.001$ cutoff (in this case $CC_{1/2} = 0.14$, $R_{meas} = 205\%$, $\langle I/\sigma \rangle_{mrgd} = 0.3$) allowed extension of the resolution from 2.4 to 2.1 Å for refinement. Authors conservatively state the resolution as

Serial femtosecond crystallography study combining single shots from 30 000–100 000 crystals per state to generate individual data sets with multiplicity of 1000–2000. The value of the multiplicity is seen in that for one data set (in their Figure S4a), merging 2000 images led to a low resolution R_{split} (similar to R_{pim}) of $\sim$30% and the extension to 64 000 images dropped that value to under 5%.

2.4 Å in the abstract, but did their analyses using the more informative 2.1 Å resolution cutoff. The authors comment on the enhanced detail visible in the extended resolution maps, and Supplementary Figure 2 contains seven examples showing subtle but real differences that improve the interpretability. The higher resolution allowed them to better define the solvation of a ribosome.

40. Wang J, Wing RA: **Diamonds in the rough: a strong case for the**
  •• **inclusion of weak-intensity X-ray diffraction data**. *Acta Crystallogr D: Biol Crystallogr* 2014, **70(Pt 5)**:1491-1497.
A compelling study showing that including weak high resolution data greatly improves the accuracy of phases obtained during phase extension using 16-fold non-crystallographic symmetry averaging. The authors also include additional analyses of their results and extensive discussion of the value of weak data. Images from this paper are included here as Figure 2b.

41. Wang J: **Estimation of the quality of refined protein crystal**
  • **structures**. *Protein Sci* 2015, **24**:661-669.
A report documenting that many PDB entries have been refined at resolutions even lower than conventional criteria would indicate and showing how this may be attractive to crystallographers because it leads to lower R/R_{free} values for any given model, even though the resulting model would generally be of worse quality than a model generated using additional higher resolution data. A novel goodness of fit measure is proposed, but not yet validated, that incorporates the number of observations (reflections) used in the refinement and so rewards the use of more data.

42. Joosten RP, Long F, Murshudov GN, Perrakis A: **The PDB_REDO**
  • **server for macromolecular structure model optimization**. *IUCrJ* 2014, **1(Pt 4)**:213-220.
Reports updates in the PDB_REDO procedures noting that these include a 'paired refinements' test. If the reflection file contains higher resolution data than the recorded resolution of the structure, PDB_REDO tests if including the additional data produces a better model using the R_{free}, the weighted R_{free}, the CC_{free}, and the free log likelihood, accepting the higher resolution limit unless two of the indicators are degraded.

43. Driggers CM, Dayal PV, Ellis HR, Karplus PA: **Crystal structure of**
  • ***Escherichia coli* SsuE: defining a general catalytic cycle for FMN reductases of the flavodoxin-like superfamily**. *Biochemistry* 2014, **53**:3509-3519.
A structure analyses for which refinement had been first completed using data sets with conventionally defined resolution limits of near 2.3 Å, but which could then be extended by ∼0.3 Å based on $CC_{1/2}$-based limits and lead to improved structures as measured by improved R/R_{free} values even at the higher resolution limit so that a paired refinement comparison was not needed to justify the extension.

44. Evans P: **Resolving some old problems in protein crystallography**. *Science* 2012, **336**:986-987.

45. Tanley SW, Diederichs K, Kroon-Batenburg LM, Schreurs AM, Helliwell JR: **Experiences with archived raw diffraction images data: capturing cisplatin after chemical conversion of carboplatin in high salt conditions for a protein crystal**. *J Synchrotron Radiat* 2013, **20(Pt 6)**:880-883.

46. Terwilliger TC: **Archiving raw crystallographic data**. *Acta Crystallogr D: Biol Crystallogr* 2014, **70(Pt 10)**:2500-2501.

47. Terwilliger TC, Bricogne G: **Continuous mutual improvement of macromolecular structure models in the PDB and of X-ray crystallographic software: the dual role of deposited experimental data**. *Acta Crystallogr D: Biol Crystallogr* 2014, **70(Pt 10)**:2533-2543.

48. Shaya D, Findeisen F, Abderemane-Ali F, Arrigoni C, Wong S,
  • Nurva SR, Loussouarn G, Minor DL Jr: **Structure of a prokaryotic sodium channel pore reveals essential gating elements and an outer ion binding site common to eukaryotic channels**. *J Mol Biol* 2014, **426**:467-483.
Used $CC_{1/2}$ ∼ 0.1 cutoff criterion to extend the resolution from 4.0 to 3.46 Å and state that 'the obvious differences in map quality reinforce the assertion that adherence to traditional metrics for defining resolution limits can result in the omission of useful diffraction data.' Images from this paper are included here as Figure 2a.